

Prevalence of *Strongyloides stercoralis* and Other Intestinal Parasite Infections in School Children in a Rural Area of Angola: A Cross-Sectional Study

María Luisa Aznar Ruiz de Alegría,^{1,2} Karen Colmenares,³ Mateu Espasa,⁴ Arancha Amor,^{5,6} Isabel Lopez,⁴ Arlette Nindia,¹ Joaquina Kanjala,¹ Domingas Guilherme,¹ Elena Sulleiro,⁴ Begoña Barriga,³ Eva Gil,¹ Fernando Salvador,^{2*} Cristina Bocanegra,² Teresa López,¹ Milagros Moreno,¹ and Israel Molina²

¹Hospital Nossa Senhora da Paz, Cubal, Angola; ²Department of Infectious Diseases, Vall d'Hebron University Hospital, PROSICS, Barcelona, Spain; ³Department of Epidemiology, Vall d'Hebron University Hospital, Barcelona, Spain; ⁴Department of Microbiology, Vall d'Hebron University Hospital, PROSICS, Barcelona, Spain; ⁵National Center of Tropical Medicine, Institute of Health Carlos III, Madrid, Spain; ⁶Mundo Sano Foundation, Buenos Aires, Argentina

Abstract. *Strongyloides stercoralis* is widely distributed in the tropics and subtropics. The aim of this study was to determine the prevalence of *S. stercoralis* and other intestinal parasites and identify the risk factors for infection with *S. stercoralis* in a rural area of Angola. A cross-sectional study was conducted in school-age children (SAC) in Cubal, Angola. A questionnaire collecting clinical and epidemiological variables was used, and two stool samples were collected. A concentration technique (Ritchie) and a technique for detection of larvae migration (Baermann) were performed. Of 230 SAC, 56.1% were female and the mean age was 9.3 years (SD 2.45). Severe malnutrition, according to body mass index (BMI)-for-age, was observed in 20.4% of the SAC, and anemia was found in 59.6%. *Strongyloides stercoralis* was observed in 28 of the 230 (12.8%) SAC. Eggs of other helminths were observed in 51 (22.2%) students: *Hymenolepis* spp. in 27 students (11.7%), hookworm in 14 (6.1%), *Schistosoma haematobium* in four (1.7%), *Enterobius vermicularis* in four (1.7%), *Ascaris lumbricoides* in three (1.3%), *Taenia* spp. in two (0.9%), and *Fasciola hepatica* in one (0.4%). Protozoa were observed in 17 (7.4%) students. Detection of *S. stercoralis* was higher using the Baermann technique versus using formol-ether (11.3 vs. 3%). Overall prevalence of *S. stercoralis* in the school population of 16 studied schools in the municipal area of Cubal was greater than 10%. This fact must be considered when designing deworming mass campaigns. The use of specific tests in larvae detection is needed to avoid overlooking this parasite.

BACKGROUND

Intestinal parasites affect more than one billion people in the world, mainly impacting impoverished populations without access to adequate water and sanitation.^{1,2} Strategies to control these infections have been focused in four species: *Ascaris lumbricoides*, *Trichuris trichiura*, *Ancylostoma duodenale*, and *Necator americanus*. *Strongyloides stercoralis*, a widely-distributed soil-transmitted helminth (STH) (Central and South America, sub-Saharan Africa, and South and Southeast Asia), has been historically neglected in the epidemiological maps.^{3,4} It has been estimated that 30–100 million people are currently infected with *S. stercoralis*, but the real prevalence is probably underestimated.^{5–9} The main reason for underestimation is poor sensitivity of most diagnostic tests.^{10–13} Infection is acquired through direct skin contact with contaminated soil and the main risk factors for the infection include walking barefoot and poor sanitation conditions. The principal difference from the rest of the STHs is its ability for autoinfection that allows it to remain in the host for long periods of time. In most cases, chronic infection is asymptomatic, although gastrointestinal and cutaneous manifestations have been documented.¹⁴ Some conditions such as the use of immunosuppressive treatment, HTLV-1 infection, and malnutrition may predispose individuals to potentially life-threatening conditions due to a massive invasion by filariform larvae, with mortality rates up to 85%.^{15–17}

The World Health Organization recommends regular treatment of 75% of preschool and SAC living in the areas where the

prevalence of STH is estimated to be more than 20% with albendazole or mebendazole; however, these drugs do not have significant activity against *S. stercoralis* in single doses.¹⁸ Consequently, there are no public health strategies to control strongyloidiasis.

In Angola, information about intestinal parasites is scarce and there are no data about the prevalence of *S. stercoralis*. To provide information about the epidemiology and identify the risk factors related to *S. stercoralis* infection, a cross-sectional study was conducted in 16 primary schools in Cubal, a rural area in South Angola.

METHODS

A cross-sectional study was conducted between January 2015 and May 2015 in the municipality of Cubal.

Study setting and population. The municipality of Cubal is situated in southwestern Angola. It has an estimated population of 322,000 people, with 151,000 (47%) of children under 15 years of age.¹⁹ It has four different districts: Cubal Center, Tumbulo, Yambala, and Capupa. The sample size calculation was based on the estimated prevalence registered in previous studies carried out in Africa.^{9,20–22} Based on the school-age population and considering a prevalence of 30%, the sample size was estimated for 378 children with an estimation error of 2.5% (CI 95%). The number of children recruited from each district was proportional to the population size of the district. We recruited children of 5–14 years in 16 schools across four districts in Cubal. Schools with the highest number of SAC in each district were chosen. Once in the school, we were given a list of all the enrolled children in alphabetical order. We then chose one in every four consecutively (i.e., the first of the list was chosen and the next three were discarded, the fifth was then chosen and so on).

* Address correspondence to Fernando Salvador, Department of Infectious Diseases, Vall d'Hebron University Hospital, PROSICS, Vall d'Hebron 119-129, Barcelona, Spain 08035. E-mail: fmsalvad@vhebron.net

Field procedures. A survey was conducted by a local health worker using a standardized questionnaire in both Portuguese and the local language of the children in the presence of their parents or guardians during daily classes. Participation in the study was voluntary and required informed consent forms to be signed by parents or legal guardians. Children who received an antiparasitary drug within the three previous months were excluded. The following information was recorded for each enrolled child: age, gender, school and district, living conditions, personal hygiene, and history of previous diseases and treatments with albendazole/mebendazole or praziquantel within the three previous months. Height and weight was obtained for all children to assess their nutritional status. Weight was recorded using an electronic scale and height was recorded using a rigid height measurement. Nutritional status was assessed and quantified using the latest WHO growth chart (2006) based on the Multicentre Growth Reference Study (MGRS) and by the BMI-for-age and height-for-age. Body mass index was calculated as weight (kg) divided by height squared (m^2). As per the WHO classification, a score of -2 to -3 indicates moderate malnutrition and a score of less than -3 indicates severe malnutrition.²³ A finger prick blood sample was obtained from each participant. Hemoglobin (Hb) concentration was measured using a HemoCueH photometer (HemoCue 201 + system, HemoCue, Angelholm, Sweden). Anemia was diagnosed when Hb was lower than 11 gr/dL—according to the Hb thresholds set by WHO.²⁴ Two stool samples from two consecutive days were obtained from all children. After the interview, each participant was given an empty, prelabeled plastic container. When possible, a filled stool container was collected the same day. If the participant could not get a sample the same day, the filled stool container was collected the next day. A second empty, prelabeled container was given the next day, and the same method was used. The samples were brought to the laboratory at ambient temperature the same day they were collected.

Laboratory procedures. Both stool samples were examined with a classic formol-ether concentration (FEC) method and Baermann method. For FEC purposes, a hazelnut-sized stool sample was mixed with 10 mL of formalin. After filtration, 2 mL of ethylic ether was added. The eluent was then centrifuged and the sediment was observed with an optic microscope for detection of both helminth eggs and protozoal cysts detection. To perform the Baermann technique, the rest of the stool sample was mixed with activated charcoal and then placed in a Petri dish over a double layer of paper towel. The sample was then covered with a single layer of paper towel. After incubating for 18 hours, the Petri dish was turned over a strainer placed over a funnel attached to a clamped rubber tube. The funnel was filled with warm water. After an hour, the content of the rubber tube was centrifuged and the sediment observed with an optic microscope for detection of *S. stercoralis* and hookworm larvae.²⁵ The larvae were differentiated based on their morphology. Each stool sample was examined for a minimum of 20 minutes by two blinded and experienced researchers. If discrepancies between researchers were present, a third laboratory technician examined the sample.

Thirty percent of the slides were randomly selected and re-examined by two qualified technicians from Vall d'Hebron University Hospital Microbiology Department, Spain, for external quality control.

Comparison between techniques. To determine which technique was better for identifying the presence of *S. stercoralis* and if the two samples yielded a higher diagnosis of *S. stercoralis* infection versus examining only one sample, we only included the children who brought two stool samples. The stool samples where we could not perform both techniques were discarded.

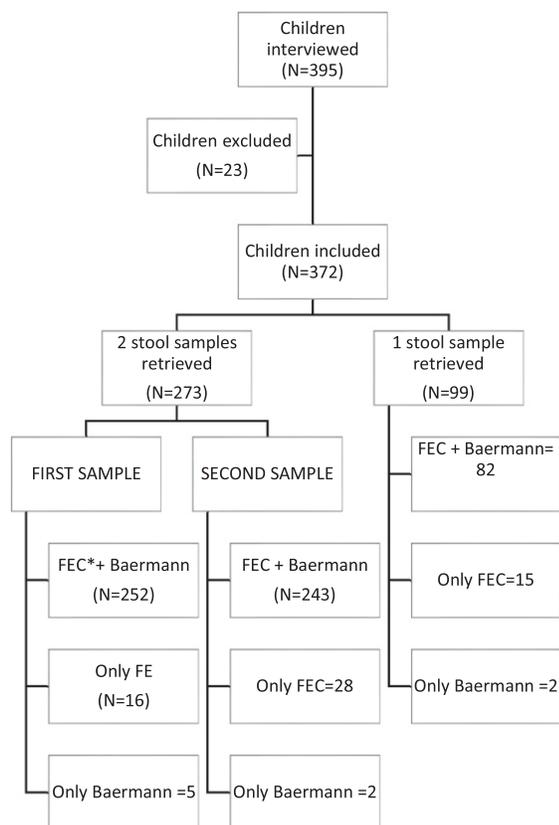
Ethics approval and consent to participate. The study was approved by the Vall d'Hebron Research Institute Ethics Committee and by the local institutions. All relevant authorities (village chiefs, school teachers, and community leaders) were informed about the purpose and procedures of the study. Written informed consent was obtained from all participants' parents or guardians, and the children participating provided oral assent. Every student diagnosed with intestinal parasites received appropriate treatment.

Statistical analysis. Data were analyzed using STATA™ version 11.0 (College Station, TX). To assess the prevalence of intestinal parasites and risk factors related with *S. stercoralis* infection, the analysis was restricted to those children who provided two stool samples and both techniques were performed in both samples. The estimated prevalence of strongyloidiasis was calculated overall and stratified by school. Absolute and relative frequencies were calculated for qualitative variables. For quantitative variables, mean and standard deviation or median and interquartile range were estimated. The normal distribution of quantitative variables was tested through the Shapiro–Wilk test. Differences of normally distributed variables between groups were evaluated using *t* tests and χ^2 tests. For the non-normally distributed variables, the differences were evaluated using Mann–Whitney *U* test. Demographics, hygiene and sanitation conditions, water source, and district were selected as potential risk factors of *S. stercoralis* infection. A final model was developed using a multivariate logistic regression analysis including the infection with *S. stercoralis* as a dependent variable (*S. stercoralis* infection yes/no as a dichotomous variable). Variables with a significance < 0.2 in the univariate analysis and variables considered to be clinically important were included as independent variables. The results have been described with odds ratios, their 95% confidence intervals (95% CI), and *P* values. *P* values < 0.05 were considered statistically significant.

RESULTS

Study sample and compliance. A total of 395 SAC were interviewed. Fifteen were excluded because of having taken antiparasitary drugs during the last three months and eight because of not providing any stool samples. Of 372 students included, 214 (57.5%) were female and mean age was 9.30 years (SD 2.41). A total of 273 students provided two stool samples, and 99 students provided a single stool sample. In some cases, owing to insufficient stool material, either FEC or Baermann techniques were not performed in all the stool samples. Figure 1 shows the distribution of the number of samples provided and the number of laboratory techniques performed in each sample. Epidemiological data, hygiene and sanitation conditions, and clinical information are shown in Table 1.

Prevalence of *S. stercoralis* and other intestinal parasites. *Strongyloides stercoralis* was observed in 28 of the 230 (12.2%)



*FEC: Formol-ether technique

FIGURE 1. Flow chart detailing compliance for stool examination.

SAC. Eggs of other helminths were observed in 51 (22.2%) students, and protozoa were observed in 17 (7.4%) SAC. Twenty-four (6.4%) children were infected with more than one intestinal parasite. Table 2 summarizes the prevalence of intestinal parasites observed.

The prevalence of *S. stercoralis* using FEC was 2.6%, whereas the prevalence rose to 11.7% using the Baermann technique. Diagnosis yield increased from 3% to 12.2% after analyzing two consecutive samples. These results are shown in Table 3.

Quality control was performed as mentioned in methods section, and we found a 70% match in the results between the local and the external technicians. False positives were observed in 8.7% of the samples, and false negatives were observed in 20.3% of the samples.

Characteristics of SAC infected with *S. stercoralis* and the risk factors associated with *S. stercoralis* infection.

When we compared the group of children infected with *S. stercoralis* with the group of children not infected with *S. stercoralis*, we observed that the presence of other intestinal parasites was more frequent in SAC infected with *S. stercoralis* than SAC not infected with *S. stercoralis* (14.3% vs. 6.4%. $P = 0.027$). Children without a latrine at home were numerically more likely to be infected with *S. stercoralis*, but this association was not statistically significant. Characteristics of infected and not infected SAC are summarized in Table 4. We found large variability between schools. We found a prevalence of more than 30% in one school from the Yambala district. In contrast, there were no SAC infected with

TABLE 1

Demographics, hygiene conditions, and clinical information of school-aged children.

Characteristics	N (%) Overall: 372
Demographics	
Gender (female)	129 (56.1)
Age, mean (SD)*	9.3 (2.4)
District	
Cubal	85 (37)
Yambala	51 (22.2)
Tumbulo	75 (32.6)
Capupa	19 (8.3)
Hygiene-sanitation conditions	
Child use to walk barefoot (yes)	151 (65.7)
Latrine at home (yes)	96 (41.7)
Water source	
River	95 (41.3)
Well	94 (40.9)
Rain	4 (1.7)
Tap water	35 (15.2)
Nutritional status	
BMI-for-age between -2 and -3 Z score	33 (14.3)
BMI-for-age < -3 Z score	47 (20.4)
Height-for-age between -2 and -3 Z score	37 (16.1)
Height-for-age < -3 Z score	21 (9.1)
Anemia (Hb < 11 gr/dL)	137 (59.6)

*SD = standard deviation; BMI = body mass index; Hb = hemoglobin.

S. stercoralis in another school from the same district and in two schools from Capupa district.

In the multivariate regression model, we did not observe any associations with *S. stercoralis* infection. These findings are summarized in Table 5.

DISCUSSION

This is the first study to assess the prevalence of *S. stercoralis* in Angola, and it shows that more than 10% of SAC are infected with this helminth in the 16 studied schools. This prevalence increased to 30% in one of the schools.

Published data from other African countries shows a heterogeneous infection rate. Cote d'Ivoire, Sudan, and Sierra Leona have reported infection rates below 5%,²⁶⁻²⁸ however, studies from Ghana and Nigeria have reported a prevalence of 17.9% and 35%, respectively.^{20,29,30} Tanzania and Ethiopia have reported prevalence similar to our study.³¹⁻³⁶ A recent study on symptomatic children under 5 years of age in Angola showed a prevalence of *S. stercoralis* of 3.5%.²² However, a comparison of these results is difficult because of the use

TABLE 2

Prevalence of helminths and protozoa

Intestinal parasites	N = 230
Intestinal helminths	
<i>Strongyloides stercoralis</i>	28 (12.2%)
<i>Hymenolepis</i> spp.	27 (11.7%)
Hookworm	14 (6.1%)
<i>Enterobius vermicularis</i>	4 (1.7%)
<i>Ascaris lumbricoides</i>	3 (1.3%)
<i>Taenia</i> spp	2 (0.9%)
<i>Fasciola hepatica</i>	1 (0.4%)
Protozoa	
<i>Giardia lamblia</i>	12 (5.2%)
<i>Balantidium coli</i>	5 (1.2%)
<i>Entamoeba histolytica/dispar</i>	3 (1.3%)

TABLE 3
Formol-ether and Baermann technique for the diagnosis of *Strongyloides stercoralis*

	Formol-ether technique	Baermann technique	Both formol-ether and Baermann techniques
One sample	5/230 (2.1%)	16/230 (6.9%)	16/230 (6.9%)
Two samples	7/230 (3%)	26/230 (11.3%)	28/230 (12.2%)

of different laboratory techniques and study population characteristics.

There are several risk factors related to *S. stercoralis* infection, including poor hygiene and sanitation conditions.⁹ Khieu et al.³⁷ observed that not using shoes or the absence of a latrine increased the incidence of *S. stercoralis* infection. In our study, none of the poor hygiene conditions previously described were associated with *S. stercoralis*. This lack of association could be explained because of the method used for data collection and verification. Family members may over-report desirable hygiene conditions which are different from the real household environment. Direct verification methods would have been necessary to avoid this possible bias.

We found large variability between schools, with one school showing a prevalence of more than 30%. We could not identify any differences in demographical data or hygiene conditions between districts or schools, so it is unknown why SAC from some schools had higher rate of infection. However, we did not assess differences in environmental factors between districts and school areas. Previous studies have reported that temperature, rainfall, and altitude are associated with the patterns of intestinal parasite infection. Differences in environmental factors between districts could explain our finding.

Detection of *S. stercoralis* was higher using the Baermann technique for larvae detection in comparison to FEC (80% of diagnostic yield increase). Most techniques used to diagnose STH based on egg detection have low sensitivity in detecting *S. stercoralis*. This is one reason for the potential global underestimation of prevalence in other studies. Several studies have shown an increase in the diagnosis of *S. stercoralis* using Baermann and Harada Mori techniques.^{38,39} However, low

and irregular elimination of *S. stercoralis* larva in feces makes it difficult to detect in a single stool sample, in comparison with several samples, as we observed in our results.⁴⁰ Molecular methods primarily based on polymerase chain reaction are also useful in diagnosing *S. stercoralis*, but we did not consider them owing to their cost and sophistication. Other diagnostic methods such as serological techniques have shown to be an alternative to diagnose *S. stercoralis*.^{9,41} However, sensitivity and specificity may differ between the different tests commercialized and the infrastructure required to perform them is more complex.

Other intestinal parasites were detected in our study, but the prevalence was lower than the data from studies performed in other areas of Angola.⁴² However, our data are similar to a recent study performed in the same area.⁴³ An interesting finding was that the prevalence of *Hymenolepis* spp. was much higher than the prevalence of other intestinal helminths. This may be because of more extended use of both albendazole and mebendazole in comparison with praziquantel because of lower cost. A recent article from Angola highlights the importance of *Hymenolepis* spp. in this country, especially in areas with poor hygiene conditions.⁴⁴

Mass drug administration (MDA) to SAC from areas with a prevalence of STH higher than 20% is the principal WHO strategy to diminish these infections.^{16,18} However, common drugs used are albendazole or mebendazole and not ivermectin which is the first-line treatment of *S. stercoralis*. Single-dose albendazole or mebendazole does not demonstrate significant activity against *S. stercoralis*. A recent review assessed the effects of ivermectin versus benzimidazoles for treating chronic *S. stercoralis* infection, and showed a higher

TABLE 4
Characteristics of school-aged children infected and not infected with *Strongyloides stercoralis*

	Presence of <i>S. stercoralis</i> (N = 28)	No presence of <i>S. stercoralis</i> (N = 202)	P
Gender (female)	15 (53.6)	114 (56.4)	0.775
Age, mean (SD)*	9.5 (2.46)	9.3 (2.45)	0.637
District			
Cubal	9 (32.14)	76 (37.6)	0.120
Yambala	11 (39.3)	40 (19.8)	-
Tumbulo	7 (25)	68 (33.7)	-
Capupa	1 (3.6)	18 (8.9)	-
Hygiene and sanitation conditions			
Child use to walk barefoot (yes)	17 (60.7)	134 (66.3)	0.557
Latrine at home (yes)	9 (32.1)	87 (43.1)	0.272
Nutritional status			
BMI-for-age < -2 Z score	8 (33.3)	72 (35.6)	0.823
Height-for-age < -2 Z score	72 (26.9)	51 (25.2)	0.854
Anemia (Hb < 11 gr/dL)	13 (54.2)	124 (61.7)	0.475
Recent medical history			
Abdominal pain	15 (53.6)	106 (52.7)	0.934
Nausea/Vomiting	5 (17.9)	39 (19.4)	0.846
Itchy skin	10 (35.7)	63 (31.5)	0.654
Antiparasitary drugs (more than 3 months before)	3 (10.7)	33 (16.3)	0.461
Other intestinal helminths	13 (46.4)	53 (26.2)	0.027

SD = standard deviation; Hb = hemoglobin; BMI = body mass index.

TABLE 5

Risk factors for *Strongyloides stercoralis* infection determined using a multivariate logistic regression model

Risk factors	OR (95 CI%)	P
District		
Yambala	2.45 (0.99–6.05)	0.053
Hygiene-sanitation conditions		
Child use to walk barefoot (yes)	0.68 (0.29–1.59)	0.374
Latrine at home (yes)	0.76 (0.30–1.96)	0.572

OR = odds ratios.

parasitological cure with ivermectin compared with albendazole.⁴⁵ Ivermectin has been extensively used in MDA for filarial worms *Onchocerca volvulus* and *Wuchereria bancrofti*. Regular ivermectin MDA for this purpose has showed an important impact on the prevalence of *S. stercoralis*.⁴⁶ In Cubal, MDA campaigns for the control of lymphatic filariasis, onchocerciasis, soil-transmitted helminthiasis control, or schistosomiasis have not been performed. Findings derived from our study suggest that strategies to control intestinal parasites should be implemented in some areas of Cubal and should include ivermectin in combination with other drugs. Beyond strongyloidiasis, the addition of ivermectin would help control other neglected tropical diseases such as lymphatic filariasis, scabies, and soil-transmitted helminthiasis.⁴⁷ Programs to control intestinal parasite infections should include interventions to promote health and nutrition, facilitate access to safe water, and educate individuals on personal hygiene measures along with MDA campaigns. The observed spatial heterogeneity should help target these campaigns to areas with higher prevalence of intestinal infections.

Both malnutrition and anemia were highly prevalent in the studied population. Studies from different areas of Angola have recently published similar data.³⁷ Although malnutrition and anemia were related to STH infection, an association between *S. stercoralis* infection and malnutrition was not observed in our study. However, these data provide evidence of a health problem in SAC that should be investigated in future studies.

Our study has several limitations. The main limitation was having a smaller sample size than expected. This was because of a considerable number of children not submitting the two stool samples requested. Children who submitted only one stool sample were excluded with the purpose of obtaining more comparable and robust data. Second, random sampling was not used in data collection, making it possible that our findings do not adequately reflect the prevalence of intestinal parasites in the area. Third, we only used fecal-based methods and not any serological techniques. Although serological tests may overestimate the prevalence of *S. Strongyloides* because of cross-reactivity with other nematode infections,⁴⁸ we believe that the combination of fecal and serological tests would increase the detection of *S. stercoralis* infection. However, the infrastructure required to perform these techniques were not available.

CONCLUSIONS

In conclusion, the prevalence of *S. stercoralis* is 12.2% in the 16 studied schools, with relevant foci more than 30%. Data from similar studies are necessary to assess the real impact of mass deworming campaigns. The use of a combination of specific tests in larvae detection and analysis of multiple stool

samples increases the probability of detection of *S. stercoralis* and should be used to avoid *S. stercoralis* misdiagnosis.

Received February 28, 2017. Accepted for publication June 16, 2017.

Published online July 31, 2017.

Acknowledgments: We acknowledge the sixth National Plan (PN) of Research + Development + Innovation (I+D+I) 2008-2011, ISCIII-General Division Networks, and Cooperative Research Centers + Collaborative Research Network on Tropical Diseases (RICET):RD12/0018/0020 and RD12/0018/0001. This article is the result of a coordinated effort with local authorities that enabled the study's success. This collaboration is tremendously important for future interventions aimed at decreasing STH infection and associated morbidity. All authors read and approved the final version of the manuscript.

Financial support: Fundação Mundo Sano. The founder of the study had no role in study design, data collection, data interpretation, or writing of the report. The authors of this article had full access to all data in the study. The corresponding author had the final responsibility for the decision to submit for publication.

Authors' addresses: María Luisa Aznar Ruiz de Alegria, Arlette Nindia, Joaquina Kanjala, Domingas Guilherme, Eva Gil, Teresa López, and Milagros Moreno, Hospital Nossa Senhora da Paz, Cubal, Angola, E-mails: maznarru@gmail.com, arlettenindia@yahoo.com, joaquinakanjala@yahoo.com, domingasguilherme@yahoo.com, evagilolivas@hotmail.com, materloga2@yahoo.es, and milamor14@yahoo.es. María Luisa Aznar Ruiz de Alegria, Fernando Salvador, Cristina Bocanegra, and Begoña Barriga, Department of Infectious Diseases, Vall d'Hebron University Hospital, PROSICS, Passeig de la Vall d'Hebron, Barcelona, Spain, E-mails: maznaru@gmail.com, fmsalvad@vhebron.net, cristinabocanegra@gmail.com, and imolina@vhebron.net. Karen Colmenares and Begoña Barriga, Department of Epidemiology, Vall d'Hebron University Hospital, Passeig de la Vall d'Hebron, Barcelona, Spain, E-mails: karenicolmenares@gmail.com and bbrubed@gmail.com. Mateu Espasa, Isabel Lopez, and Elena Sulleiro, Department of Microbiology, Vall d'Hebron University Hospital, PROSICS, Passeig de la Vall d'Hebron, Barcelona, Spain, E-mails: mespasa@vhebron.net, maibel56@gmail.com, and esulleir@vhebron.net. Arancha Amor, National Center of Tropical Medicine, Institute of Health Carlos III, Madrid, Spain, and Mundo Sano Foundation, Castellana 13, Madrid, Spain, E-mail: aranchazu@gmail.com.

REFERENCES

- Pullan RL, Smith JL, Jasrasaria R, Brooker SJ, 2014. Global numbers of infection and disease burden of soil transmitted helminth infections in 2010. *Parasit Vectors* 7: 37.
- de Silva NR, Brooker S, Hotez PJ, Montresor A, Engels D, Savioli L, 2003. Soil-transmitted helminth infections: updating the global picture. *Trends Parasitol* 19: 547–551.
- Olsen A, et al., 2009. Strongyloidiasis—the most neglected of the neglected tropical diseases? *Trans R Soc Trop Med Hyg* 103: 967–972.
- Krolewiecki AJ, et al., 2013. A public health response against *Strongyloides stercoralis*: time to look at soil-transmitted helminthiasis in full. *PLoS Negl Trop Dis* 7: e2165.
- Genta RM, 1989. Global prevalence of strongyloidiasis: critical review with epidemiologic insights into the prevention of disseminated disease. *Rev Infect Dis* 11: 755–767.
- Utzinger J, et al., 2012. Neglected tropical diseases: diagnosis, clinical management, treatment and control. *Swiss Med Wkly* 142: w13727.
- Jorgensen T, Montresor A, Savioli L, 1996. Effectively controlling strongyloidiasis. *Parasitol Today* 12: 164.
- Puthiyakunnon S, et al., 2014. Strongyloidiasis—an insight into its global prevalence and management. *PLoS Negl Trop Dis* 8: e3018.
- Schär F, et al., 2013. *Strongyloides stercoralis*: global distribution and risk factors. *PLoS Negl Trop Dis* 7: e2288.
- Bisoffi Z, et al., 2014. Diagnostic accuracy of five serologic tests for *Strongyloides stercoralis* infection. *PLoS Negl Trop Dis* 8: e2640.

11. Knopp S, et al., 2014. Diagnostic accuracy of Kato-Katz, FLOTAC, Baermann, and PCR methods for the detection of light-intensity hookworm and *Strongyloides stercoralis* infections in Tanzania. *Am J Trop Med Hyg* 90: 535–545.
12. Sultana Y, Jeoffreys N, Watts MR, Gilbert GL, Lee R, 2013. Real-time polymerase chain reaction for detection of *Strongyloides stercoralis* in stool. *Am J Trop Med Hyg* 88: 1048–1051.
13. Levenhagen MA, Costa-Cruz JM, 2014. Update on immunologic and molecular diagnosis of human strongyloidiasis. *Acta Trop* 135: 33–43.
14. Khieu V, Srey S, Schär F, Muth S, Marti H, Odermatt P, 2013. *Strongyloides stercoralis* is a cause of abdominal pain, diarrhea and urticaria in rural Cambodia. *BMC Res Notes* 6: 200.
15. Mejia R, Nutman TB, 2012. Screening, prevention, and treatment for hyperinfection syndrome and disseminated infections caused by *Strongyloides stercoralis*. *Curr Opin Infect Dis* 25: 458–463.
16. Keiser PB, Nutman TB, 2004. *Strongyloides stercoralis* in the immunocompromised population. *Clin Microbiol Rev* 17: 208–217.
17. Carvalho EM, Da Fonseca Porto A, 2004. Epidemiological and clinical interaction between HTLV-1 and *Strongyloides stercoralis*. *Parasite Immunol* 26: 487–497.
18. WHO, 2006. *Preventive Chemotherapy in Human Helminthiasis: Coordinated Use of Anthelmintic Drugs in Control Interventions: A Manual for Health Professionals and Programme Managers*. Geneva, Switzerland: World Health Organization.
19. Administração Municipal do Cubal EdALE, 2009. Perfil do Município do CUBAL, Província de Benguela 2009.
20. Agi PI, 1997. Comparative helminth infections of man in two rural communities of the Niger Delta, Nigeria. *West Afr J Med* 16: 232–236.
21. Becker SL, et al., 2015. Real-time PCR for detection of *Strongyloides stercoralis* in human stool samples from Côte d'Ivoire: diagnostic accuracy, inter-laboratory comparison and patterns of hookworm co-infection. *Acta Trop* 150: 210–217.
22. Gasparinho C, et al., 2016. Etiology of diarrhea in children younger than 5 years attending the Bengo general hospital in Angola. *Pediatr Infect Dis J* 35: e28–e34.
23. Group WMGRS, 2006. WHO child growth standards based on length/height, weight and age. *Acta Paediatr Suppl* 450: 76–85.
24. WHO, 2001. *Iron Deficiency Anaemia: Assessment, Prevention and Control. A Guide for Programme Managers*. Geneva, Switzerland: World Health Organization.
25. Garcia LS. *Diagnostic Medical Parasitology*. Washington, DC: American Society for Microbiology, 1–172.
26. Gbakima AA, Sahr F, 1995. Intestinal parasitic infections among rural farming communities in eastern Sierra Leone. *Afr J Med Med Sci* 24: 195–200.
27. Yapi YG, Briet OJ, Vounatsou P, 2006. Prevalence of geohelminths in savana and forest areas of Côte d'Ivoire. *West Afr J Med* 25: 124–125.
28. Magambo JK, Zeyhle E, Wachira TM, 1998. Prevalence of intestinal parasites among children in southern Sudan. *East Afr Med J* 75: 288–290.
29. Verweij JJ, et al., 2009. Molecular diagnosis of *Strongyloides stercoralis* in faecal samples using real-time PCR. *Trans R Soc Trop Med Hyg* 103: 342–346.
30. Dada-Adegbola HO, Bakare RA, 2004. Strongyloidiasis in children five years and below. *West Afr J Med* 23: 194–197.
31. Knopp S, et al., 2008. Diagnosis of soil-transmitted helminths in the era of preventive chemotherapy: effect of multiple stool sampling and use of different diagnostic techniques. *PLoS Negl Trop Dis* 2: e331.
32. Fontanet AL, et al., 2000. Epidemiology of infections with intestinal parasites and human immunodeficiency virus (HIV) among sugar-estate residents in Ethiopia. *Ann Trop Med Parasitol* 94: 269–278.
33. Assefa S, Erko B, Medhin G, Assefa Z, Shimelis T, 2009. Intestinal parasitic infections in relation to HIV/AIDS status, diarrhea and CD4 T-cell count. *BMC Infect Dis* 9: 155.
34. Knopp S, et al., 2009. Changing patterns of soil-transmitted helminthiasis in Zanzibar in the context of national helminth control programs. *Am J Trop Med Hyg* 81: 1071–1078.
35. Getaneh A, Medhin G, Shimelis T, 2010. Cryptosporidium and *Strongyloides stercoralis* infections among people with and without HIV infection and efficiency of diagnostic methods for *Strongyloides* in Yirgalem Hospital, southern Ethiopia. *BMC Res Notes* 3: 90.
36. Kawai K, Saathoff E, Antelman G, Msamanga G, Fawzi WW, 2009. Geophagy (Soil-eating) in relation to Anemia and Helminth infection among HIV-infected pregnant women in Tanzania. *Am J Trop Med Hyg* 80: 36–43.
37. Khieu V, et al., 2014. High prevalence and spatial distribution of *Strongyloides stercoralis* in rural Cambodia. *PLoS Negl Trop Dis* 8: e2854.
38. Siddiqui AA, Berk SL, 2001. Diagnosis of *Strongyloides stercoralis* infection. *Clin Infect Dis* 33: 1040–1047.
39. Sato Y, Kobayashi J, Toma H, Shiroma Y, 1995. Efficacy of stool examination for detection of *Strongyloides* infection. *Am J Trop Med Hyg* 53: 248–250.
40. Nielsen PB, Mojon M, 1987. Improved diagnosis of *Strongyloides stercoralis* by seven consecutive stool specimens. *Zentralbl Bakteriol Mikrobiol Hyg [A]* 263: 616–618.
41. Luvira V, et al., 2016. Comparative diagnosis of strongyloidiasis in immunocompromised patients. *Am J Trop Med Hyg* 95: 401–404.
42. Sousa-Figueiredo JC, et al., 2012. Epidemiology of malaria, schistosomiasis, geohelminths, anemia and malnutrition in the context of a demographic surveillance system in northern Angola. *PLoS One* 7: e33189.
43. Bocanegra C, et al., 2015. Epidemiology of schistosomiasis and usefulness of indirect diagnostic tests in school-age children in Cubal, Central Angola. *PLoS Negl Trop Dis* 9: e0004055.
44. Soares Magalhães RJ, et al., 2013. Extending helminth control beyond STH and schistosomiasis: the case of human hymenolepiasis. *PLoS Negl Trop Dis* 7: e2321.
45. Henriquez-Camacho C, Gotuzzo E, Echevarria J, White AC Jr, Terashima A, Samalvides F, Pérez-Molina JA, Plana MN, 2016. Ivermectin versus albendazole or thiabendazole for *Strongyloides stercoralis* infection. *Cochrane Database Syst Rev* 1: CD007745
46. Anselmi M, et al., 2015. Mass administration of ivermectin for the elimination of onchocerciasis significantly reduced and maintained low the prevalence of *Strongyloides stercoralis* in Esmeraldas, Ecuador. *PLoS Negl Trop Dis* 9: e0004150.
47. Albonico M, et al., 2016. StrongNet: an international network to improve diagnostics and access to treatment for strongyloidiasis control. *PLoS Negl Trop Dis* 10: e0004898.
48. Requena-Méndez A, Chiodini P, Bisoffi Z, Buonfrate D, Gotuzzo E, Muñoz J, 2013. The laboratory diagnosis and follow up of strongyloidiasis: a systematic review. *PLoS Negl Trop Dis* 7: e2002.