

available at www.sciencedirect.comjournal homepage: eunoncology.europeanurology.com

European Association of Urology



Development and Independent Validation of a Prognostic Gene Expression Signature Based on RB1, PTEN, and TP53 in Metastatic Hormone-sensitive Prostate Cancer Patients

Natalia Jiménez^{a,1}, Marta Garcia de Herreros^{a,b,1}, Òscar Reig^{a,b,c,d}, Mercedes Marín-Aguilera^{a,b}, Caterina Aversa^{a,b,c}, Laura Ferrer-Mileo^{a,b,c}, Samuel García-Esteve^{a,d}, Leonardo Rodríguez-Carunchio^{c,e}, Isabel Trias^{c,e}, Albert Font^f, Alejo Rodríguez-Vida^g, Miguel Ángel Climent^h, Sara Crosⁱ, Isabel Chirivella^j, Montserrat Domènech^k, Mariona Figols^k, Joan Carles^l, Cristina Suárez^l, Daniel Herrero Rivera^m, Enrique González-Billalabeitiaⁿ, Claudia Cívico^o, Núria Sala-González^p, Vicenç Ruiz de Porrás^q, Maria J. Ribal^r, Aleix Prat^{a,b,d}, Begoña Mellado^{a,b,c,d,*,1}

^a Translational Genomics and Targeted Therapeutics in Solid Tumors Lab, Fundació de Recerca Clínic Barcelona – Institut d'Investigacions Biomèdiques August Pi i Sunyer (FRCB-IDIBAPS), Barcelona, Spain; ^b Medical Oncology Department, Hospital Clínic, Barcelona, Spain; ^c Uro-Oncology Unit, Hospital Clínic, University of Barcelona, Barcelona, Spain; ^d Department of Medicine, University of Barcelona, Barcelona, Spain; ^e Department of Pathology, Hospital Clínic, Barcelona, Spain; ^f Medical Oncology Department, Institut Català d'Oncologia, Hospital Germans Trias i Pujol, Badalona, Spain; ^g Medical Oncology Department, Institut Hospital del Mar d'Investigacions Mèdiques (IMIM), Hospital del Mar, Barcelona, Spain; ^h Medical Oncology Service, Instituto Valenciano de Oncología (IVO), Valencia, Spain; ⁱ Medical Oncology Department, Hospital General de Granollers, Barcelona, Spain; ^j Oncology Department, Hospital Clínic Universitario de Valencia, Valencia, Spain; ^k Medical Oncology Department, Fundació Althaia, Xarxa Assistencial Universitària de Manresa, Spain; ^l Vall d'Hebron Institute of Oncology, Vall d'Hebron University Hospital, Barcelona, Spain; ^m Medical Oncology Department, Hospital Virgen del Rocío, Sevilla, Spain; ⁿ Medical Oncology Department, University Hospital, 12 de Octubre, Madrid, Spain; ^o Department of Hematology and Medical Oncology, Hospital Universitario Morales Meseguer, IMIB-Universidad de Murcia, Murcia, Spain; ^p Department of Medical Oncology, Institut Català d'Oncologia, Girona, Spain; ^q Badalona Applied Research Group in Oncology (B-ARGO), Institut Català d'Oncologia – Germans Trias i Pujol Research Institute, Badalona, Spain; ^r Department of Urology, Hospital Clínic, Barcelona, Spain

Article info

Article history:

Received 31 October 2023

Received in Revised form

30 November 2023

Accepted 29 December 2023

Associate Editor:

Elena Castro

Keywords:

Hormone-sensitive prostate cancer

Tumor suppressor genes

Biomarkers

Abstract

Background: Androgen deprivation therapy (ADT) with docetaxel (D) and/or antiandrogen receptor therapies (ARTs) are the standard therapies in metastatic hormone-sensitive prostate cancer (mHSPC). Alterations in the tumor suppressor genes (TSGs) *RB1*, *PTEN*, and *TP53* are associated with an aggressive evolution and treatment resistance in castration-resistant prostate cancer (CRPC).

Objective: To study the clinical implications of TSG mRNA expression in mHSPC patients.

Design, setting, and participants: This is a multicenter retrospective biomarker study in mHSPC patients. TSG_{low} status was defined when two or more out of the three TSGs presented low RNA expression by nCounter in formalin-fixed paraffin-embedded samples and TSG_{wt} for the remaining cases. The microarray data from the CHARTED trial were analyzed as an independent validation cohort.

Outcome measurements and statistical analysis: Molecular data were correlated with CRPC-free survival (CRPC-FS) and overall survival (OS) by the Kaplan-Meier method and multivariate Cox analysis.

¹ These authors contributed equally to this work.

* Corresponding author. Medical Oncology Department, Hospital Clínic of Barcelona, Villarroel 170, 08036 Barcelona, Spain. Tel. +34 93 227 54 00.

E-mail address: bmellado@clinic.cat (B. Mellado).

<https://doi.org/10.1016/j.euo.2023.12.012>

2588-9311/© 2024 The Author(s). Published by Elsevier B.V. on behalf of European Association of Urology. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Please cite this article as: N. Jiménez, M. Garcia de Herreros, Ò. Reig et al., Development and Independent Validation of a Prognostic Gene Expression Signature Based on *RB1*, *PTEN*, and *TP53* in Metastatic Hormone-sensitive Prostate Cancer Patients, *Eur Urol Oncol* (2024), <https://doi.org/10.1016/j.euo.2023.12.012>

Androgen deprivation therapy
Docetaxel
CHAARTED trial

Results and limitations: A total of 226 patients were included, of whom 218 were eligible: 93 were treated with ADT and 125 with ADT + D; 75.7% presented de novo stage IV and 67.9% high-volume disease. TSG_{low} (19.2%) was independently correlated with shorter CRPC-FS (hazard ratio [HR] 1.8, $p = 0.002$) and OS (HR 2, $p = 0.002$). In the CHAARTED trial, TSG_{low} was independently correlated with lower CRPC-FS (HR 2.2, $p = 0.02$); no differences in clinical outcomes according to treatment were observed in TSG_{low} patients, while a significant benefit was observed for ADT + D in the TSG_{wt} group for CRPC-FS (HR 0.4, $p < 0.001$) and OS (HR 0.4, $p = 0.001$). However, no interaction was observed between TSG signature and treatment in either series. Study limitations are the retrospective design, small sample size, and lack of inclusion of patients treated with ADT + ART.

Conclusions: TSG_{low} expression correlates with adverse outcomes in patients with mHSPC. The investigation of new therapeutic strategies in these patients is warranted.

Patient summary: The low RNA expression of tumor suppressor genes in the tumors is correlated with adverse outcomes in patients with metastatic hormone-sensitive prostate cancer.

© 2024 The Author(s). Published by Elsevier B.V. on behalf of European Association of Urology. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Prostate cancer is ranked as second in cancer incidence and the fifth cause of cancer death in men [1]. Androgen deprivation therapy (ADT) with docetaxel (D) or antiandrogen receptor therapies (ARTs) are the standard upfront treatments in metastatic hormone-sensitive prostate cancer (mHSPC) [2–8]. Moreover, the addition of ARTs to ADT + D, with either darolutamide or abiraterone, has also shown survival benefits in patients with synchronous mHSPC regardless of the risk and volume of disease [9–11]. However, treatment selection remains a challenge and biomarkers are needed.

Alterations in the tumor suppressor genes (TSGs) *RB1*, *PTEN*, and *TP53* have been associated with the development of aggressive variant prostate cancer and neuroendocrine (NE) dedifferentiation in castration-resistant prostate cancer (CRPC) [12]. These variants usually appear after antiandrogen therapies, are defined by distinct clinical features and androgen receptor (AR)-independent progression, and are associated with reduced response to conventional therapies, poor prognosis, and more sensitivity to platinum [13].

The role of TSG alterations in mHSPC is less well defined. In this context, exome mutations at least in one gene can be detected in about 30% of patients [14,15]. Moreover, genomic alterations in two or more TSGs in men with HSPC and CRPC are associated with poor clinical outcomes [13].

The transcriptional profile of primary tumors may determine a distinct clinical evolution and treatment benefit of mHSPC patients. A subanalysis of the CHAARTED clinical trial [2], which compared ADT + D versus ADT alone treatments in mHSPC, identified that patients with a luminal B molecular subtype benefited from the addition of D to ADT, in contrast to the basal subtype. More recently, a molecular analysis of the ADT-treated patient cohorts with or without abiraterone from the STAMPEDE trial identified several prognostic transcriptional signatures, as low mRNA expression of *PTEN* or *TP53* [16].

In a prior study in mHSPC patients treated with ADT + D, we found that the low TSG expression signature correlated

with lower overall survival (OS). Moreover, patients with lowest tertile expression of at least two TSGs presented shorter CRPC-free survival (CRPC-FS) and OS [17].

In the current study, we define and further validate a TSG_{low} signature in a larger series of mHSPC patients with extended follow-up, and explore its potential value for treatment selection. Additionally, we independently validate these results through an in silico analysis of molecular data from patients included in the phase 3 CHAARTED trial [18].

2. Patients and methods

Complete details are given in the [Supplementary material](#).

2.1. Design, patients, and samples

We present a multicenter retrospective biomarker study in patients with mHSPC from ten hospitals in Spain. The key inclusion criteria were as described previously [17]. The study was conducted according to the principles of the Declaration of Helsinki, and it was approved by the institutional ethics committees of all participating centers. Informed consent was obtained from all patients. Treatment for mHSPC was ADT alone (ie, luteinizing hormone-releasing hormone analogs) or ADT in combination with D (75 mg/m² every 21 d for six cycles).

The primary endpoint was to correlate TSG mRNA expression with CRPC-FS. The secondary endpoints were to correlate TSG mRNA expression and OS, to study the correlation between loss-of-function exome mutations with mRNA expression and immunohistochemistry (IHC), and to explore the impact of the determination of TSGs through different techniques on clinical outcomes.

2.2. Gene expression panel design

We configured a gene expression nCounter panel (Nanos-tring Technologies, Seattle, WA, USA) of 184 genes [17]. Here, we present the data focused on the TSG signature and also explore the expression of the full-length *AR*.

2.3. Bioinformatics and statistical analysis

Tertiles were applied to transformed (z score) nCounter gene expression data from an exploratory series to establish the cutoff for *RB1*, *PTEN*, and *TP53* expression, and categorize the samples as high-, mid-, or low-expression groups for each gene. These cutoffs were then applied to the transformed (z score) gene expression data from the other cohorts described in the Results section and the microarray data from CHARTED trial patients [18]. Low term was assigned for the lower tertile, and *wt* for the mid and high tertiles of each gene. TSG_{low} was considered when two or more out of the three TSGs presented low expression and TSG_{wt} for the remaining cases.

CRPC-FS and OS, calculated from the date of start of ADT to the time of developing CRPC, and to the time of death or last follow-up visit, respectively, were analyzed by the Kaplan-Meier method and compared by log-rank test. CRPC-FS definition, treatment response criteria, and progressive disease definitions followed the Prostate Cancer Working Group 2 criteria [19]. Univariate and multivariate analyses of variables of interest were performed by a Cox regression analysis. Analyses were performed with R software (version 3.6.3; R Foundation for Statistical Computing, Vienna, Austria).

3. Results

3.1. Patients, samples, and TSG expression signature

A total of 226 patients were enrolled in the study: 218 were eligible and eight were excluded due to insufficient tumor sample ($n = 4$) or RNA quantity ($n = 4$). Of the eligible

patients, 125 were treated with ADT + D and 93 with ADT alone (Table 1). Most formalin-fixed paraffin-embedded (FFPE) samples were obtained from the primary tumor (93.1%). The median follow-up was 46.3 (range 6.7–223.5) mo. As shown in Figure 1A, 24.4%, 30.5%, and 23.9% of patients were considered RB1_{low}, PTEN_{low}, and TP53_{low}, respectively. According to our criteria, 19.2% of patients were TSG_{low}. Moreover, we explored whether there were differences in *AR* expression between TSG_{low} and TSG_{wt} groups, observing that *AR* mRNA levels were lower in TSG_{low} ($p = 0.002$; Fig. 1B).

Overall, there were no differences in clinical characteristics between TSG_{low} and TSG_{wt} (Supplementary Tables 1–6). Regarding individual TSG RNA levels, low *PTEN* expression correlated with de novo stage IV disease in the global series ($p = 0.022$) and the ADT + D cohort ($p = 0.004$), and with visceral metastases in the ADT + D cohort ($p = 0.049$). Moreover, *RB1* ($p = 0.017$) and *TP53* ($p = 0.047$) expression correlated with visceral metastases in the ADT cohort (Supplementary Fig. 1–3).

3.2. Comparison between TSG determination by nCounter and other techniques

In 60 patients, TSG mRNA was determined by both nCounter and RNA-Seq, observing a high correlation of mRNA levels of each gene by both techniques (Supplementary Fig. 4).

A targeted TSG mutation analysis was performed in 54 patients treated with ADT + D. Mutations in at least one TSG were present in 30 patients (55.6%); *RB1* was mutated in 11 (20.4%), *PTEN* in 18 (33.3%), and *TP53* in 19 (35.2%) patients, whereas 14 (25.9%) presented mutations in more

Table 1 – Characteristics of patients from the global cohort^a

	Global cohort	ADT + D cohort	ADT cohort	<i>p</i> value
Patients, <i>n</i> (%)	218	125 (57.3)	93 (42.7) ^b	
Age (yr)				
Median (range)	66.4 (46.3–84.6)	66.6 (46.3–83.4)	66.1 (51–84.6)	0.467
Tumor origin, <i>n</i> (%)				
Primary	203 (93.1)	117 (93.6)	86 (92.5)	0.791
Metastatic	15 (6.9)	8 (6.4)	7 (7.5)	
Stage at diagnosis, <i>n</i> (%)				
<IV	42 (19.3)	9 (7.2)	33 (35.5)	<0.001
IV	165 (75.7)	116 (92.8)	49 (52.7)	
NA	11 (5)	–	11 (11.8)	
Gleason sum at diagnosis, <i>n</i> (%)				
≤7	53 (24.3)	22 (17.6)	31 (33.3)	0.004
≥8	158 (72.5)	102 (81.6)	56 (60.2)	
NA	7 (3.2)	1 (0.8)	6 (6.5)	
Presence of visceral metastases, <i>n</i> (%)				
Yes	33 (15.1)	25 (20)	8 (8.6)	0.034
No	181 (83)	100 (80)	81 (87.1)	
NA	4 (1.9)	–	4 (4.3)	
Disease volume, <i>n</i> (%)				
High	148 (67.9)	97 (77.6)	51 (54.8)	0.002
Low	65 (29.8)	27 (21.6)	38 (40.9)	
NA	5 (2.3)	1 (0.8)	4 (4.3)	
ECOG performance status score, <i>n</i> (%)				
0	93 (42.7)	54 (43.2)	39 (41.9)	0.777
1 or 2	114 (52.3)	69 (55.2)	45 (48.4)	
NA	11 (5.0)	2 (1.6)	9 (9.7)	

ADT = androgen deprivation therapy; D = docetaxel; ECOG = Eastern Cooperative Oncology Group; *n* = number of cases; NA = not available.

^a The *p* values are based on Fisher exact test and Wilcoxon Mann-Whitney *U* test for categorical and continuous variables, respectively. Significant *p* values ($p < 0.05$) are bold indicated.

^b Five patients were excluded from survival analysis due to lack of complete follow-up data.

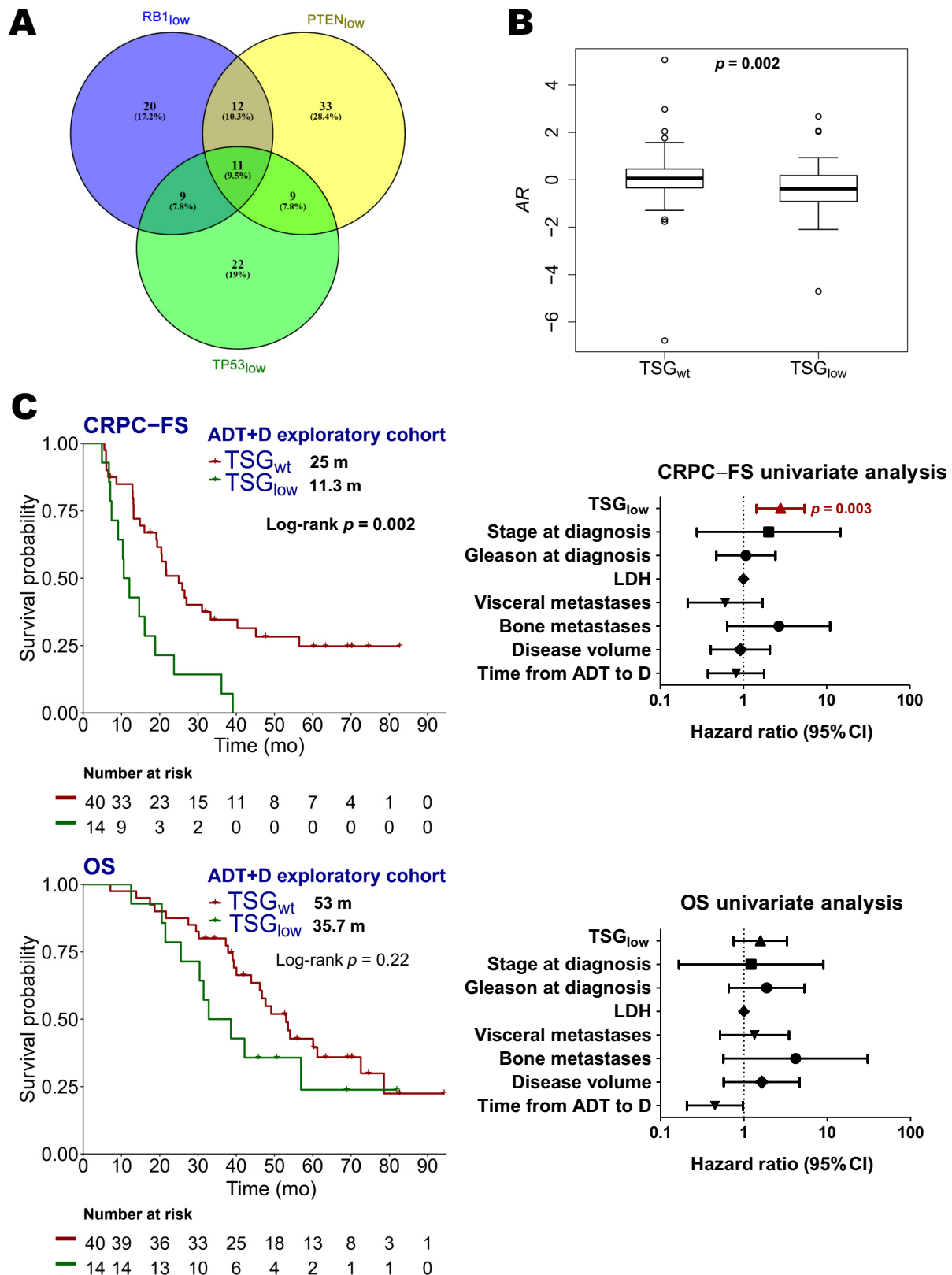


Fig. 1 – (A) Venn diagram of the $RB1_{low}$, $PTEN_{low}$, and $TP53_{low}$ patients with complete follow-up in the global (ADT \pm D) cohort. (B) Boxplot of androgen receptor (AR) RNA expression levels (z score) according to TSG expression in the global (ADT \pm D) cohort (Wilcoxon test; p value). (C) Kaplan-Meier curves representing CRPC-FS and OS according to TSG expression and forest plots representing the univariate analysis in the ADT + D exploratory cohort. ADT = androgen deprivation therapy; CI = confidence interval; CRPC = castration-resistant prostate cancer; CRPC-FS = CRPC-free survival; D = docetaxel; LDH = lactate dehydrogenase; m: median months; OS = overall survival; TSG = tumor suppressor gene. Significant p values ($p < 0.05$) are bold indicated.

than two TSGs and four (7.4%) in all three genes. Most of the pathogenic variants were missense (46.9%) or nonsense mutations (46.7%). Details of pathogenic mutations found in each TSG are shown in [Supplementary Table 7](#).

A significant correlation was observed between the presence of mutations in *PTEN* and low *PTEN* RNA levels ($p = 0.026$; [Supplementary Fig. 5A](#)). The mutational status of none of the individual TSGs was associated with clinical outcomes ([Supplementary Fig. 5B and 5C](#)). The presence of two or more TSG mutations (TSG_{mut}) was correlated with shorter CRPC-FS (hazard ratio [HR] 2, 95% confidence interval [CI] 1.1–4, $p = 0.036$; [Supplementary Fig. 5D](#)).

IHC was carried out in tumor samples from 73 patients from the ADT + D cohort with available tumor for IHC. Finally, 48 (65.8%) samples were assessable for RB1, 52 (71.2%) for *PTEN*, and 56 (76.7%) for *TP53*. Thirty-eight (79.2%) samples presented alterations in RB1, 26 (50%) in *PTEN*, and 28 (50%) in *TP53*. Altered IHC for RB1 and *PTEN* correlated significantly with low levels of RNA expression ($p = 0.007$ and $p < 0.001$, respectively; [Supplementary Fig. 6A](#)). The alteration by IHC of none of the individual TSGs correlated with clinical outcomes, nor having two or more altered TSGs (in 34 [66.7%] patients; [Supplementary Fig. 6B–D](#)).

3.3. TSG mRNA expression signature in an exploratory cohort

The series of 54 patients from the ADT + D cohort, with both mutational and nCounter expression data, was considered the exploratory cohort ([Supplementary Table 8](#)). In this cohort, TSG_{low} correlated with shorter CRPC-FS (HR 2.8, 95% CI 1.4–5.4, $p = 0.002$; [Fig. 1C](#)). Moreover, the model that included the TSG assessed by mRNA expression (Akaike Information Criterion [AIC] score: 275.8) fitted better than the one that included their mutational status (AIC: 279.8).

3.4. TSG mRNA expression signature validation in patients treated with ADT + D

Internal validation of the results was performed in the additional 71 patients treated with ADT + D ([Supplementary Table 8](#)), where TSG_{low} correlated with lower CRPC-FS (HR 2.4, 95% CI 1.1–5.3, $p = 0.033$) and OS (HR 3.2, 95% CI 1.4–7.3, $p = 0.006$). Moreover, TSG_{low} was independently associated with shorter CRPC-FS (HR 4.1, 95% CI 1.6–10.4, $p = 0.003$) and OS (HR 3.7, 95% CI 1.6–8.7, $p = 0.003$; [Fig. 2](#)).

3.5. TSG mRNA expression signature in patients treated with ADT alone

The established cutoffs were also analyzed in a cohort of 93 patients treated with ADT alone ([Table 1](#)). In this cohort, TSG_{low} was not associated with either CRPC-FS (HR 1.4, 95% CI 0.8–2.3, $p = 0.23$) or OS (HR 1.6, 95% CI 0.9–2.7, $p = 0.091$; [Fig. 3](#)).

3.6. Exploring TSG mRNA expression as a predictor of treatment benefit

To explore whether TSG mRNA expression was a predictor of treatment benefit, we analyzed together the patients treated with ADT + D and those treated with ADT (global cohort;

[Table 1](#)). TSG_{low} patients had shorter CRPC-FS (HR 1.9, 95% CI 1.3–2.7, $p = 0.001$) and OS (HR 1.8, 95% CI 1.2–2.7, $p = 0.002$) than TSG_{wt} patients. Moreover, TSG_{low} correlated independently with shorter CRPC-FS (HR 1.8, 95% CI 1.3–2.7, $p = 0.002$) and OS (HR 2, 95% CI 1.3–3.1, $p = 0.002$; [Fig. 4](#)). However, no interaction between TSG expression and treatment was observed regarding CRPC-FS ($p = 0.11$) or OS ($p = 0.45$).

3.7. Independent series validation

In the microarray data from the CHARTED trial [18], 27.5% of the patients were classified as TSG_{low} patients. In the multivariate analysis, TSG_{low} was independently correlated with shorter CRPC-FS (HR 2.2, 95% CI 1.1–4.3, $p = 0.02$; [Fig. 5A](#)).

Analyzing TSG_{low} and TSG_{wt} populations separately according to treatment, we found that there were no significant differences in CRPC-FS ($p = 0.3$) or OS ($p = 0.5$) between TSG_{low} patients treated with ADT + D or ADT alone. Moreover, TSG_{wt} patients treated with ADT + D had the longest CRPC-FS (HR 0.4, 95% CI 0.2–0.6, $p < 0.001$) and OS (HR 0.4, 95% CI 0.3–0.7, $p = 0.001$), compared with ADT-treated patients. However, as observed in our series, no interaction between the TSG expression and treatment was observed regarding CRPC-FS ($p = 0.116$) or OS ($p = 0.051$; [Fig. 5B and 5C](#)).

3.8. Individual assessment of TSG

In the global cohort, $RB1_{low}$ (HR 1.6, 95% CI 1.2–2.2, $p = 0.006$) and $PTEN_{low}$ (HR 1.8, 95% CI 1.3–2.5, $p < 0.001$) correlated with CRPC-FS. Moreover, $RB1_{low}$ (HR 1.6, 95% CI 1.1–2.2, $p = 0.018$), $PTEN_{low}$ (HR 1.7, 95% CI 1.2–2.3, $p = 0.003$), and $TP53_{low}$ (HR 1.5, 95% CI 1.1–2.2, $p = 0.023$) correlated with OS ([Supplementary Fig. 7](#)). In the multivariate analysis, $RB1_{low}$ correlated with CRPC-FS (HR 1.5, 95% CI 1–2.1, $p = 0.03$), $PTEN_{low}$ correlated with CRPC-FS (HR 1.6, 95% CI 1.2–2.3, $p = 0.003$) and OS (HR 1.5, 95% CI 1.1–2.2, $p = 0.018$), and $TP53_{low}$ correlated with OS (HR 1.6, 95% CI 1.1–2.4, $p = 0.013$; [Supplementary Fig. 8](#)).

The multivariate analysis including TSG_{low} was the best accurate model for both CRPC-FS and OS compared with those that included the low expression from an individual gene ([Supplementary Table 9](#)).

4. Discussion

In this study, we show that the mRNA expression of the TSG_{low} signature (low expression of two or more of the TSGs *RB1*, *PTEN*, and *TP53*) is independently associated with lower CRPC-FS and OS in mHSPC patients. The prognostic value of the TSG signature was validated independently in the molecular dataset from patients included in the CHARTED trial [2,18]. Besides, we found that the lower expression of any of the individual genes was also independently associated with an adverse prognosis, although the TSG_{low} signature was a better model for CRPC-FS and OS prediction. We also explored whether the TSG signature could be useful to predict treatment benefit. We found that in the CHARTED series, when analyzing TSG_{low} and TSG_{wt}

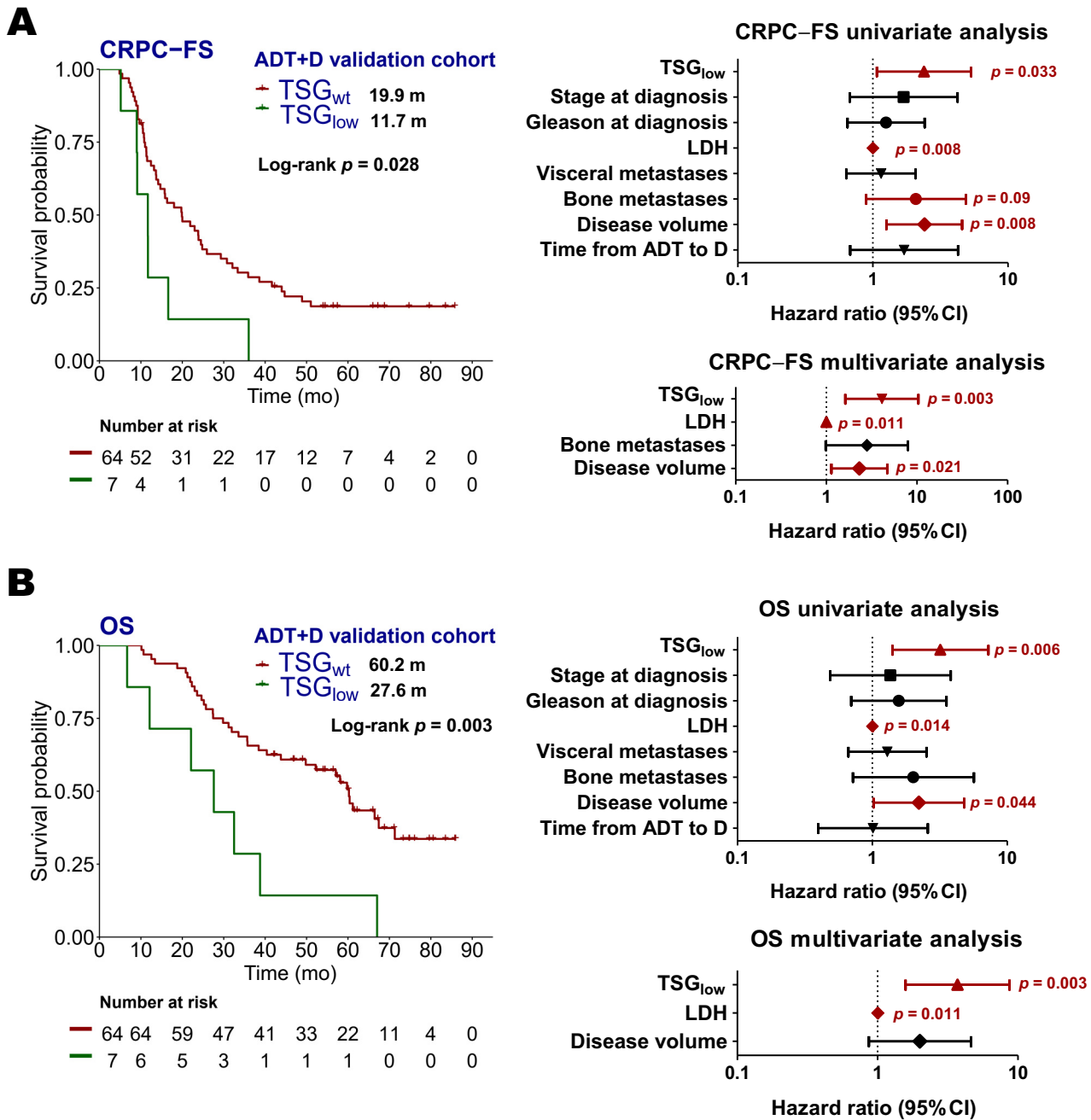


Fig. 2 – Kaplan-Meier curves representing (A) CRPC-FS and (B) OS according to TSG expression and forest plots representing the univariate and multivariate analyses in the ADT + D validation cohort. ADT = androgen deprivation therapy; CI = confidence interval; CRPC = castration-resistant prostate cancer; CRPC-FS = CRPC-free survival; D = docetaxel; LDH = lactate dehydrogenase; m = median months; OS = overall survival; TSG = tumor suppressor gene. Significant p values ($p < 0.05$) are bold indicated.

populations separately according to treatment, there were no significant differences in CRPC-FS and OS between TSG_{low} patients treated with ADT alone or ADT + D. Moreover, TSG_{wt} patients treated with ADT + D presented longer CRPC-FS and OS than those treated with ADT alone. However, no interaction was observed between the TSG signature and treatment in either our study or the CHARTED series. Thus, we may not conclude that TSG_{low} patients do not benefit from adding D to ADT.

As one of the established standards of care for mHSPC is ADT + ART, it will be relevant to test the TSG signature in

patients receiving this treatment strategy, as well as in those treated with ADT + ART + D [9–11], in order to elucidate whether they would benefit from D addition.

While most of the previous studies have focused on studying TSG genomic alterations or IHC protein expression, just a few of them have analyzed TSG RNA expression. Both IHC and next-generation sequencing (NGS) have been proved to be able to correlate TSG alterations with clinical outcomes, but they have not been compared rigorously [12,15]. One study conducted in tumor-derived xenografts, which studied TSG alterations by IHC, RNA expression, and

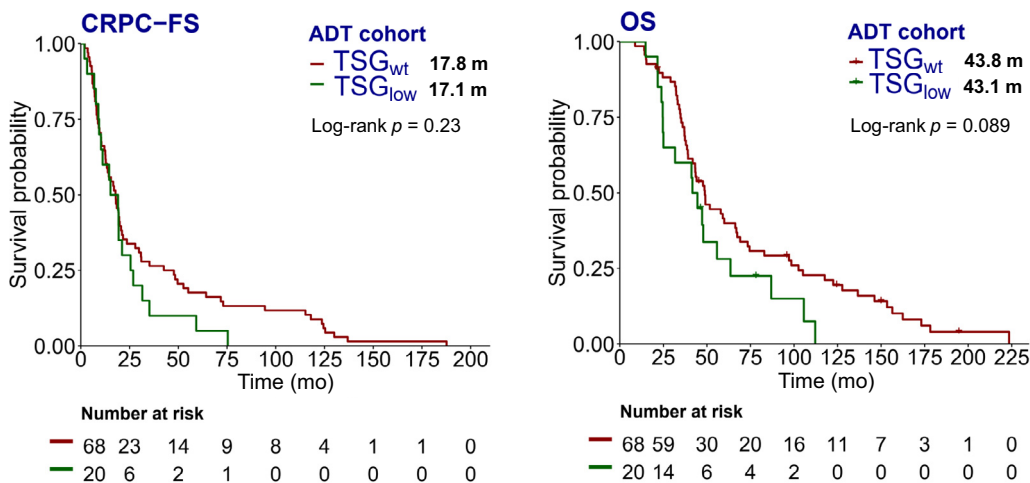


Fig. 3 – Kaplan-Meier curves representing CRPC-FS and OS according to TSG expression in the ADT cohort. ADT = androgen deprivation therapy; CRPC = castration-resistant prostate cancer; CRPC-FS = CRPC-free survival; m = median months; OS = overall survival; TSG = tumor suppressor gene.

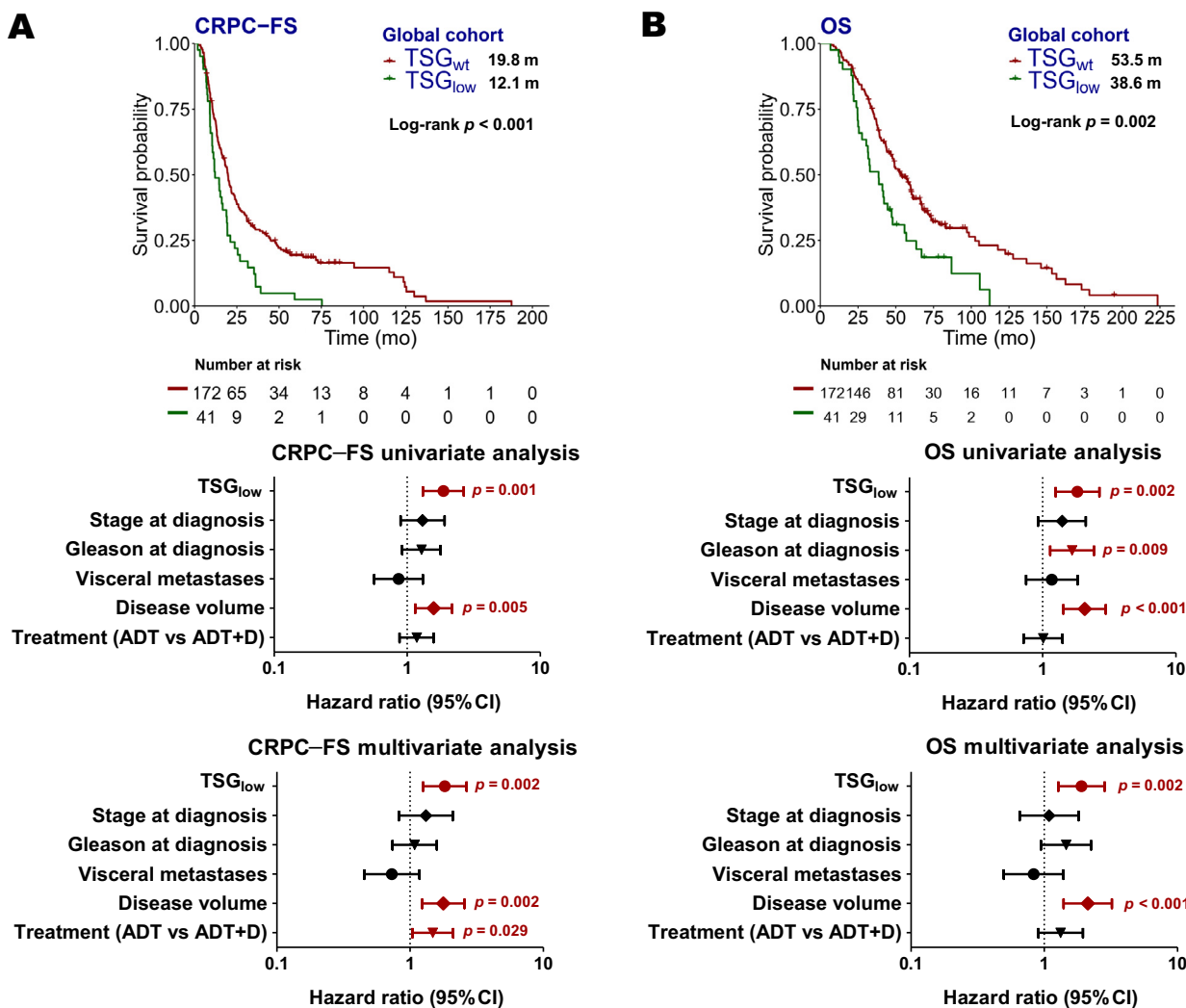


Fig. 4 – Kaplan-Meier curves representing (A) CRPC-free survival (CRPC-FS) and (B) overall survival (OS) according to TSG expression and forest plots representing the univariate and multivariate analyses in the global (ADT ± Docetaxel [D]) cohort. ADT = androgen deprivation therapy; CI = confidence interval; CRPC = castration-resistant prostate cancer; m = median months; TSG = tumor suppressor gene. Significant *p* values (*p* < 0.05) are bold indicated.

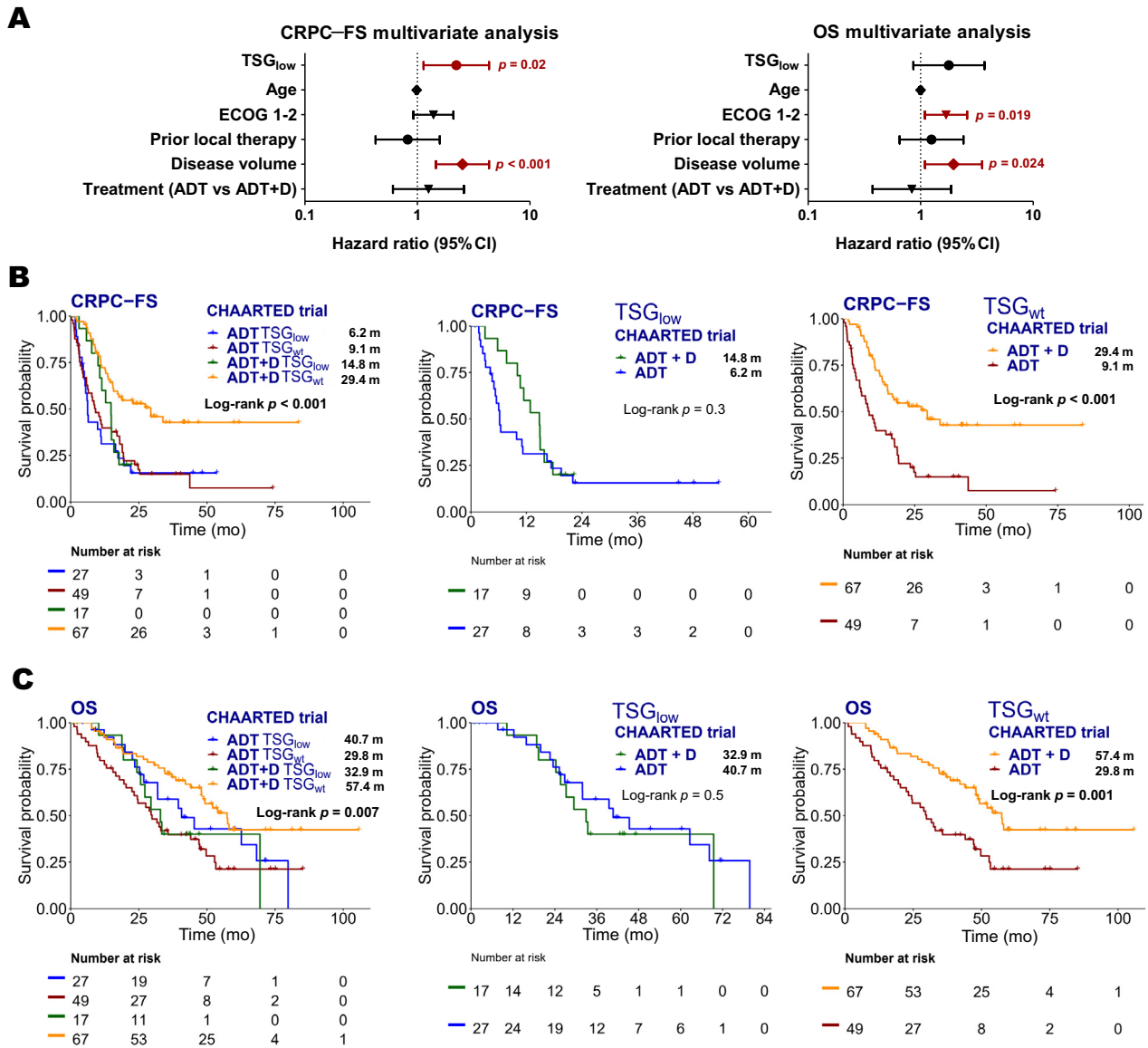


Fig. 5 – (A) Forest plots representing the multivariate analysis of TSG expression of microarray data from the CHAARTED trial for CRPC-free survival (CRPC-FS) and overall survival (OS). Kaplan-Meier curves representing (B) CRPC-FS and (C) OS according to TSG expression in the CHAARTED trial segregated by treatment: ADT + docetaxel (D) arm and ADT arm. ADT = androgen deprivation therapy; CI = confidence interval; CRPC = castration-resistant prostate cancer; ECOG = Eastern Cooperative Oncology Group; m = median months; TSG = tumor suppressor gene. Significant p values ($p < 0.05$) are bold indicated.

DNA sequencing, found a good genotype-to-phenotype correlation [20]. Here, we have studied TSG alterations by DNA, RNA, and IHC in a subset of patients. We have observed a correlation between mRNA expression of *PTEN* and *PTEN* mutations, and between mRNA and IHC expression of *PTEN* and *RB1*. For *TP53*, we could not find a strong association between IHC patterns, mutations, and RNA expression. As prostate tumor tissues show low basal expression of *TP53*, IHC cannot detect *TP53* loss that results from nonsense, frameshift, or indel alterations that may also lead to low RNA expression [21,22]. Moreover, there is a lack of standardized criteria for TSG determination by IHC in prostate cancer.

Focusing on DNA-RNA discordance, cases with lower RNA expression without any genetic alteration may be explained by post-transcriptional alterations, changes in methylation

patterns, or interactions with noncoding RNA that can affect mRNA expression without the presence of a mutation [23]. Moreover, some genetic variants such as copy number alterations or large deletions are often not detected by NGS conventional assays. In our study, mRNA expression of TSG was a better outcome predictor than genomic alterations.

TP53, *PTEN*, and *RB1* are recurrently altered in CRPC. In this context, the presence of two or more TSG alterations (mainly defined by genomic loss or mutations or altered protein expression by IHC) is associated with aggressive clinical features, resistance to conventional therapies, aggressive evolution, and NE dedifferentiation [12,24–26]. A gene expression signature reflecting *TP53/RB1* loss is associated with diminished responses to AR antagonists and reduced survival [27].

It is known that molecular alterations of primary prostate cancer may differ from those of CRPC. Mateo et al. [15] studied genomic aberrations in matched hormone-naïve and CRPC biopsies from 61 patients who developed mCRPC, and found differences in *TP53*, *RB1*, and *PI3K/AKT* mutational status between same-patient samples. Furthermore, cell plasticity-related changes that occur as a result of ARTs [24,28] may not be present in treatment-naïve primary tumors. Notably, in one study, 40% of *TP53/RB1*-deficient tumors were classified as AR-active adenocarcinomas; therefore, NE differentiation is not a necessary consequence of *TP53/RB1* inactivation [26]. Similarly, in a prior study in mHSPC, we did not find a correlation between TSG and NE markers mRNA expression [17]. Thus, the absence of NE markers expression in mHSPC does not exclude the presence of TSG alterations. Moreover, we found in the present study that lower TSG expression correlated with lower AR expression. Thus, TSG alterations in noncastrated tumors may preclude the development of NE dedifferentiation and androgen-independent progression during CRPC progression [29].

Several studies addressed the clinical implications of TSG genomic alterations in mHSPC patients. In a large massively parallel targeted sequencing study, where patients with altered TSG were defined by harboring any copy number loss or deleterious mutation of one or more TSGs, authors found that patients with prostate tumors with compound TSG mutations had poorer outcomes [13]. A meta-analysis and systematic review of 11 studies including 1682 mHSPC patients found that high-volume and de novo mHSPC were enriched with *TP53* alterations [30]. We found in the present study that lower *PTEN* levels correlated with de novo mHSPC in the global and ADT + D series and the presence of visceral metastases in the ADT + D series, and that lower *RB1* and *TP53* expression correlated with visceral metastases in the ADT series. However, analyzing together, we did not find differences in clinical characteristics between TSG_{low} and the rest of the patients, and notably, the TSG_{low} signature was an independent adverse prognostic factor for CRPC-FS and OS [17]. This may suggest that this molecular signature may be more accurate than clinical characteristics in predicting the outcome in mHSPC patients.

5. Conclusions

In conclusion, our study shows the adverse prognostic factor of the TSG_{low} signature in mHSPC patients. The investigation of this signature in patients receiving ADT + ART or the triple therapy with ADT, ART, and D may be of interest in order to determine the benefit of D addition according to the TSG status. Overall, the adverse clinical implications of having TSG alterations support the investigation of new therapeutic strategies in metastatic prostate cancer patients with these molecular alterations.

Author contributions: Begoña Mellado had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Study concept and design: Mellado, Jiménez, García de Herreros, Ò. Reig, M. Marín-Aguilera.

Acquisition of data: Mellado, Jiménez, García de Herreros, Reig, Marín-Aguilera, Aversa, Ferrer-Mileo, García-Esteve, Rodríguez-Carunchio, Trias, Font, Rodríguez-Vida, Climent, Cros, Chirivella, Domènech, Figols, Carles, Suárez, Herrero Rivera, González-Billalabeitia, Cívico, Sala-González.

Analysis and interpretation of data: Mellado, Jiménez, García de Herreros, Reig, Marín-Aguilera, Aversa, Ferrer-Mileo, García-Esteve, Rodríguez-Carunchio, Trias.

Drafting of the manuscript: Mellado, Jiménez, García de Herreros, Reig. **Critical revision of the manuscript for important intellectual content:** All authors.

Statistical analysis: Mellado, Jiménez, García de Herreros, Reig, Marín-Aguilera.

Obtaining funding: Mellado.

Administrative, technical, or material support: Mellado, Jiménez, García de Herreros, Reig, Marín-Aguilera, Aversa, Ferrer-Mileo, García-Esteve, Rodríguez-Carunchio, Trias, Font, Ruiz de Porras, Ribal, Prat.

Supervision: Mellado.

Other: None.

Financial disclosures: Begoña Mellado certifies that all conflicts of interest, including specific financial interests and relationships and affiliations relevant to the subject matter or materials discussed in the manuscript (eg, employment/affiliation, grants or funding, consultancies, honoraria, stock ownership or options, expert testimony, royalties, or patents filed, received, or pending), are the following: The authors have provided the following conflicts to disclose (which may not be related to the subject matter of this manuscript): Begoña Mellado: research funding from Janssen, Roche, Bayer, and Pfizer; speakers' bureau for Roche, Sanofi, Janssen, Astellas, Pfizer, Novartis, and Bristol-Myers Squibb; and travel and accommodation expenses from Janssen and Pfizer. Óscar Reig: consulting or advisory role for BMS, EISAI, and Ipsen; and travel and accommodation expenses from Ipsen and Pfizer. Caterina Aversa: speaker honoraria from BMS, Janssen, and Pfizer, and travel expenses from Janssen. Laura Ferrer-Mileo: speaker honoraria and travel accommodation expenses from Pfizer and Kyowa kirin, and research funding from Roche. Albert Font: research funding from AstraZeneca; consulting or advisory role for Janssen, Astellas, and Bayer; and travel and accommodation expenses from Janssen. Alejo Rodríguez-Vida: research funding from Takeda, MSD, and Pfizer; consulting or advisory role for Astellas, Bayer, BMS, MSD, Janssen, Roche, Pfizer, and Clovis; and honoraria or travel expenses from Pfizer, MSD, Astellas, BMS, Janssen, AstraZeneca, Roche, Bayer, and Sanofi Aventis. Miguel Ángel Climent: consulting or advisory role for BMS, MSD, Bayer, EUNSA, Pfizer, Roche, Janssen, Pierre Fabre, and Ipsen, and travel and accommodation expenses from Janssen, Astellas, Roche, Ipsen, and MSD. Sara Cros: research funding from Pfizer and Janssen; consulting or advisory role for GSK; and speakers' bureau for GSK, BMS, and Merck. Isabel Chirivella: advisory boards for Pfizer, EISAI, and BMS. Montserrat Domènech: consulting or advisory and/or speakers' bureau for Sanofi Aventis, Bristol-Myers Squibb, and Pfizer, and travel and accommodations expenses from Sanofi and Lilly. Mariona Figols: speakers' bureau for Pfizer, Ipsen, and Astellas, and travel and accommodation expenses from Merck. Joan Carles: consulting for Astellas Pharma, AstraZeneca, Bayer, Bristol-Myers Squibb, Johnson & Johnson, MSD Oncology, Novartis, Pfizer, Roche, and Sanofi, and speakers' bureau for Bayer, Asofarma, Astellas, and Janssen. Cristina Suárez: consulting or advisory and/or speakers' bureau for Astellas, Bayer, BMS, Roche, Ipsen, Merck Sharp & Dohme, Novartis, Pfizer, and Sanofi. Daniel Herrero Rivera: honoraria from Pfizer and IQVIA, and travel and accommodations expenses from GILEAD sciences. Enrique

González-Billalabeitia: advisory board for Astellas, Janssen, Sanofi, Astra-Zeneca, and Bayer, and speakers' bureau for Astellas, Bayer, and Janssen. Núria Sala-González: advisory board for Pfizer, Bristol Myers Squibb, and Roche, and speakers' bureau for Ipsen and Astellas Pharma. Maria J. Ribal: speaker honoraria from Ipsen, Janssen, and Astellas. Aleix Prat: advisory and consulting fees from Roche, Pfizer, Novartis, Amgen, BMS, Puma, Oncolytics Biotech, MSD, Guardant Health, Peptomyc, and Lilly; lecture fees from Roche, Pfizer, Novartis, Amgen, BMS, Daiichi Sankyo, and Nanostring technologies; institutional financial interests from Boehringer, Novartis, Roche, Nanostring technologies, Sysmex Europe GmbH, Medica Scientia Innovation Research, SL, Celgene, Astellas, and Pfizer; leadership role in Reveal Genomics, SL; and a patent PCT/EP2016/080056. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Funding/Support and role of the sponsor: This work was supported by Instituto de Salud Carlos III-Subdirección General de Evaluación y Fomento de la Investigación (PI18/714) and cofunded by the European Union. Institutional funding from CERCA Programme/Generalitat de Catalunya is gratefully acknowledged. This work was funded by a grant from Janssen-Pharmaceuticals (212082PCR4056) and an Astellas General Research Grant (ID: 71843877). Òscar Reig is awarded with a "Ayudas SEOM de Intensificación para Investigadores Jovenes" from the Spanish Society of Medical Oncology (SEOM). This work was developed at the Centro Esther Koplowitz and CELLEX, Barcelona, Spain.

Acknowledgments: The authors thank Esther Barnadas for her kind organization of sample collection from Hospital Clínic and her excellent technical assistance with FFPE tumor sections. We also want to thank Dr. Christopher J. Sweeney (Dana Farber/Harvard Cancer Center) for his kind suggestions and comments on this work. We want to acknowledge Parc de Salut MAR Biobank (MARBiobanc; RD09/0076/00036, PT17/0015/0011) and IGT-PHUGTP Biobank (PT13/0010/0009, PT17/0015/0045) integrated in the Spanish National Biobanks Network and Tumor Bank Network of Catalonia, the BioBank FIVO (PT17/0015/0051) integrated in the Spanish National Biobanks Network and in the Valencian Biobanking Network, and the Andalusian Public Health System Biobank (SSPA Biobank) for their collaboration in providing samples. The authors are also deeply indebted to all patients who agreed to be involved in the study. The authors acknowledge ECOG-ACRIN and NCTN/NCORP for sharing clinical and gene expression data. This manuscript utilized data from datasets NCT00309985-D3 and NCT00309985-D5 from the NCTN/NCORP Data Archive of the National Cancer Institute's (NCI's) National Clinical Trials Network (NCTN). Data were originally collected from clinical trial NCT00309985 (E3805 study) entitled "Androgen ablation therapy with or without chemotherapy in treating patients with metastatic prostate cancer (CHAARTED)." We want to also thank the E3805 investigators Dr. Christopher J. Sweeney (Dana Farber/Harvard Cancer Center), Dr. Robert S. DiPaola (University of Kentucky College of Medicine), and Yu-Hui Chen (Dana-Farber Cancer Institute/ECOG-ACRIN Biostatistics Center) for sharing the data (financial grants U10CA180820, U10CA180794, and UG1CA233180). Taxotere was provided by Sanofi. All analyses and conclusions in this manuscript are the sole responsibility of the authors and do not necessarily reflect the opinions or official views of the National Institutes of Health, clinical trial investigators, the NCTN, the NCORP, or the NCI.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.euo.2023.12.012>.

References

- [1] Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2021;71:209–49.
- [2] Sweeney CJ, Chen Y-H, Carducci M, et al. Chemohormonal therapy in metastatic hormone-sensitive prostate cancer. *N Engl J Med* 2015;373:737–46.
- [3] Gravis G, Boher J-M, Chen Y-H, et al. Burden of metastatic castrate naive prostate cancer patients, to identify men more likely to benefit from early docetaxel: further analyses of CHAARTED and GETUG-AFU15 studies. *Eur Urol* 2018;73:847–55.
- [4] James ND, Sydes MR, Clarke NW, et al. Addition of docetaxel, zoledronic acid, or both to first-line long-term hormone therapy in prostate cancer (STAMPEDE): survival results from an adaptive, multiarm, multistage, platform randomised controlled trial. *Lancet* 2016;387:1163–77.
- [5] Fizazi K, Tran N, Fein L, et al. Abiraterone plus prednisone in metastatic, castration-sensitive prostate cancer. *N Engl J Med* 2017;377:352–60.
- [6] James ND, de Bono JS, Spears MR, et al. Abiraterone for prostate cancer not previously treated with hormone therapy. *N Engl J Med* 2017;377:338–51.
- [7] Davis ID, Martin AJ, Stockler MR, et al. Enzalutamide with standard first-line therapy in metastatic prostate cancer. *N Engl J Med* 2019;381:121–31.
- [8] Chi KN, Agarwal N, Bjartell A, et al. Apalutamide for metastatic, castration-sensitive prostate cancer. *N Engl J Med* 2019;381:13–24.
- [9] Fizazi K, Foulon S, Carles J, et al. Abiraterone plus prednisone added to androgen deprivation therapy and docetaxel in de novo metastatic castration-sensitive prostate cancer (PEACE-1): a multicentre, open-label, randomised, phase 3 study with a 2 × 2 factorial design. *Lancet* 2022;399:1695–707.
- [10] Smith MR, Hussain M, Saad F, et al. Darolutamide and survival in metastatic, hormone-sensitive prostate cancer. *N Engl J Med* 2022;386:1132–42.
- [11] Sweeney CJ, Martin AJ, Stockler MR, et al. Testosterone suppression plus enzalutamide versus testosterone suppression plus standard antiandrogen therapy for metastatic hormone-sensitive prostate cancer (ENZAMET): an international, open-label, randomised, phase 3 trial. *Lancet Oncol* 2023;24:323–34.
- [12] Aparicio AM, Shen L, Tapia ELN, et al. Combined tumor suppressor defects characterize clinically defined aggressive variant prostate cancers. *Clin Cancer Res* 2016;22:1520–30.
- [13] Hamid AA, Gray KP, Shaw G, et al. Compound genomic alterations of TP53, PTEN, and RB1 tumor suppressors in localized and metastatic prostate cancer. *Eur Urol* 2019;76:89–97.
- [14] Velez MG, Kosiorek HE, Egan JB, et al. Differential impact of tumor suppressor gene (TP53, PTEN, RB1) alterations and treatment outcomes in metastatic, hormone-sensitive prostate cancer. *Prostate Cancer Prostatic Dis* 2022;25:479–83.
- [15] Mateo J, Seed G, Bertan C, et al. Genomics of lethal prostate cancer at diagnosis and castration resistance. *J Clin Invest* 2020;130:1743–51.
- [16] Attard G, Parry M, Grist E, et al. Clinical testing of transcriptome-wide expression profiles in high-risk localized and metastatic prostate cancer starting androgen deprivation therapy: an ancillary study of the STAMPEDE abiraterone phase 3 trial. *Res Sq* 2023;rs.3.rs-2488586. <https://doi.org/10.21203/rs.3.rs-2488586/v1>.
- [17] Jiménez N, Reig Ó, Marín-Aguilera M, et al. Transcriptional profile associated with clinical outcomes in metastatic hormone-sensitive prostate cancer treated with androgen deprivation and docetaxel. *Cancers* 2022;14:4757.
- [18] Hamid AA, Huang H-C, Wang V, et al. Transcriptional profiling of primary prostate tumor in metastatic hormone-sensitive prostate cancer and association with clinical outcomes: correlative analysis of the E3805 CHAARTED trial. *Ann Oncol* 2021;32:1157–66.
- [19] Scher HI, Halabi S, Tannock I, et al. Design and end points of clinical trials for patients with progressive prostate cancer and castrate levels of testosterone: recommendations of the Prostate Cancer Clinical Trials Working Group. *J Clin Oncol* 2008;26:1148–59.
- [20] Soundararajan R, Viscuse P, Pilie P, et al. Genotype-to-phenotype associations in the aggressive variant prostate cancer molecular profile (AVPC-m) components. *Cancers* 2022;14:3233.
- [21] Navone NM, Troncso P, Pisters LL, et al. p53 Protein accumulation and gene mutation in the progression of human prostate carcinoma. *J Natl Cancer Inst* 1993;85:1657–69.

- [22] Tan H-L, Sood A, Rahimi HA, et al. Rb loss is characteristic of prostatic small cell neuroendocrine carcinoma. *Clin Cancer Res* 2014;20: 890–903.
- [23] Jia P, Zhao Z. Impacts of somatic mutations on gene expression: an association perspective. *Brief Bioinform* 2017;18:413–25.
- [24] Beltran H, Prandi D, Mosquera JM, et al. Divergent clonal evolution of castration-resistant neuroendocrine prostate cancer. *Nat Med* 2016;22:298–305.
- [25] Abida W, Cyrta J, Heller G, et al. Genomic correlates of clinical outcome in advanced prostate cancer. *PNAS* 2019;116:11428–36.
- [26] Nyquist MD, Corella A, Coleman I, et al. Combined TP53 and RB1 loss promotes prostate cancer resistance to a spectrum of therapeutics and confers vulnerability to replication stress. *Cell Rep* 2020;31: 107669.
- [27] Ku SY, Rosario S, Wang Y, et al. Rb1 and Trp53 cooperate to suppress prostate cancer lineage plasticity, metastasis, and antiandrogen resistance. *Science* 2017;355:78–83.
- [28] Davies AH, Beltran H, Zoubeidi A. Cellular plasticity and the neuroendocrine phenotype in prostate cancer. *Nat Rev Urol* 2018;15:271–86.
- [29] Conteduca V, Ku S-Y, Fernandez L, et al. Circulating tumor cell heterogeneity in neuroendocrine prostate cancer by single cell copy number analysis. *NPJ Precis Oncol* 2021;5:76.
- [30] Van der Eecken K, Vanwelkenhuyzen J, Deek MP, et al. Tissue- and blood-derived genomic biomarkers for metastatic hormone-sensitive prostate cancer: a systematic review. *Eur Urol Oncol* 2021;4:914–23.